DOI: 10.31285/AGRO.29.1587 ISSN 2730-5066

Identification of the pseudoautosomal region of the X chromosome in sheep and sex prediction using the ARS-UI_Ramb_v2.0 genome assembly

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Editor

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Received 13 Oct 2024 Accepted 07 May 2025 Published 13 Jun 2025

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Abstract

Sex determination through genotyping is an efficient tool for verifying recorded sex in sheep. Since males possess only one X chromosome, heterozygous genotypes should not be present in the non-pseudoautosomal region (nPAR) of the X chromosome. Therefore, determining the boundaries of the pseudoautosomal region (PAB) is necessary for a reliable sex prediction. Although it is recommended to use SNPs from both the X and Y chromosomes, the SNP chip we used did not contain Y chromosome markers. This study aimed to determine the pseudoautosomal region (PAR) on the X chromosome in sheep using the ARS-UI_Ramb_v2.0 genome assembly and test a method for sex prediction based solely on X chromosome SNPs. The training dataset was composed of 210 sheep from various breeds and crossbreeds, genotyped with the Ovine Infinium® HD SNP BeadChip to determine the PAR region. A validation dataset of 229 sheep was used to assess the accuracy of sex determination using the OvineSNP50 BeadChip. After quality control, the PAR region was found to span from 0 to 7.24 Mb and was identified by SNPs that exhibited high heterozygosity rates in males. The proposed approach, that uses only X chromosome SNPs, achieved a sex prediction accuracy of 94% with precision rates of 99% for males and 89% for females. This study showed that it is possible to predict sex in sheep using only X chromosome nPAR SNPs. This is the first study that has used this approach specifically using the ARS-UI Ramb v2.0 genome assembly.

Keywords: ARS-I_Ramb_v2.0, pseudoautosomal region, sex prediction, X chromosome

Identificación de la región pseudoautosómica del cromosoma X en ovinos y predicción del sexo mediante el ensamblado ARS-UI_Ramb_v2.0

Resumen

La determinación de sexo mediante genotipado es eficaz para verificar el sexo declarado en el pedigrí. Como los machos poseen un solo cromosoma X, no se deberían observar genotipos heterocigotos en la región no pseudoautosómica (nPAR). Por lo tanto, es fundamental establecer claramente los límites de la región pseudoautosómica (PAB) para una correcta identificación del sexo. Aunque se recomienda utilizar SNP de los cromosomas X e Y, el chip de genotipa-





do utilizado en este estudio no contenía marcadores del cromosoma Y. El objetivo de este trabajo fue identificar la región pseudoautosómica (PAR) en el cromosoma X en ovinos utilizando el ensamblado ARS-UI_Ramb_v2.0 y evaluar un método para la predicción de sexo basado únicamente en los SNP del cromosoma X. Para definir la región PAR, se utilizó una población de 210 ovinos, genotipados con el Ovine Infinium® HD SNP BeadChip, y para la validación, 229 ovinos, genotipados con el OvineSNP50 BeadChip. Después del control de calidad, se observó que la región PAR se extendía desde 0 a 7,24 Mb, identificada mediante SNP que exhibieron altas tasas de heterocigosidad en machos. El enfoque propuesto, que utiliza únicamente los SNP del cromosoma X, alcanzó una exactitud de 94% en la predicción de sexo, con una precisión de 99% en machos y de 89% en hembras. Este estudio demostró que es posible predecir el sexo en ovinos utilizando únicamente los SNP de la región nPAR del cromosoma X. Este es el primer trabajo en aplicar este método utilizando el ensamblado ARS-UI_Ramb_v2.0.

Palabras clave: ARS-I_Ramb_v2.0, región pseudoautosómica, predicción del sexo, cromosoma X

Identificação da região pseudoautossômica do cromossomo X em ovinos e predição de sexo usando a montagem do genoma ARS-UI_Ramb_v2.0

Resumo

A determinação do sexo por genotipagem é uma ferramenta eficiente para verificar o sexo registrado. Como os machos possuem apenas um cromossomo X, os genótipos heterozigotos não devem estar presentes na região não pseudoautossômica (nPAR) do cromossomo X. Portanto, determinar os limites da região pseudoautossômica (PAB) é necessário para uma predição confiável do sexo. Embora seja recomendado usar SNP dos cromossomos X e Y, o chip SNP que usamos não continha marcadores do cromossomo Y. Este estudo teve como objetivo determinar a região pseudoautossômica (PAR) no cromossomo X em ovinos usando o genoma ARS-UI_Ramb_v2.0 e testar um método para predição de sexo baseado apenas em SNP do cromossomo X. Para definição da região PAR foi utilizada uma população de treinamento de 210 ovinos genotipados com o Ovine Infinium® HD SNP BeadChip e para validação, 229 ovinos genotipados com o OvineSNP50 BeadChip. Após controle de qualidade, observou-se que a região PAR se estendia de 0 a 7,24 Mb, identificada por SNP que exibiam altas taxas de heterozigosidade em machos. A abordagem proposta, que usa apenas SNPs do cromossomo X, atingiu uma exatidão de predição de sexo de 94%, com taxas de precisão de 99% para machos e 89% para fêmeas. Este estudo mostrou que a predição de sexo em ovinos pode ser feita usando apenas SNP da região nPAR do cromossomo X. Este é o primeiro estudo que usou esta abordagem especificamente usando o genoma ARS-UI_Ramb_v2.0.

Palavras-chave: ARS-I_Ramb_v2.0, região pseudoautossômica, predição de sexo, cromossomo X

1. Introduction

In a genotyping pipeline, sex determination through genotyping serves as an effective quality control (QC) measure that offers a straightforward QC check to ensure that the genotyped DNA corresponds to the reported animal⁽¹⁾. Gender classification mistakes due to incorrect sample identification, data entry mistakes, or genotyping errors can cause delays in genetic progress of breeding programs⁽²⁾.

The sex-calling algorithm that is used in Affymetrix chips is one of the ways of determining sex and is available in the proprietary Axiom Analysis SuiteTM software. This algorithm is named as cn-probe-chrXYratio_sex. This method of sex determination is based on the ratio of the average probe intensity of non-polymorphic probes on the Y chromosome to those on the X chromosome, excluding the copy number probes within the PAR regions of both the X and Y chromosomes. Another approach is through X chromosome heterozygosity⁽³⁾. Sheep [*Ovis aries*] have 26 pairs of autosomal chromosomes and two sex chromosomes: X and Y. Females, with two X chromosomes, display heterozygosity, whereas males, with one X and one Y chromosome, are hemizygous for the X chromosome⁽³⁾. However, an exception is the pseudoautosomal region (PAR), a region of sequence



homology between the X chromosome and Y chromosome that is involved in pairing, recombination, and segregation during meiosis, where males can also be heterozygous⁽⁴⁾⁽⁵⁾. It is important to identify the PAR region for the correct determination or confirmation of sex from X chromosome genotypes⁽²⁾. Although sex prediction using SNPs from both X and Y chromosomes is optimal, not all commercial SNP chips contain Y chromosome markers. This is because assembling the mammalian Y chromosome has been quite difficult, as it contains a lot of repetitive DNA sequences that complicate sequencing and assembly⁽⁶⁾⁽⁷⁾. In the past, it was rather challenging to obtain high-quality assembly of the Y chromosome in the Bovidae family. However, a recent study by Olagunju and others⁽⁷⁾ reported contiguous, high-quality Y chromosome assemblies in both sheep and cattle, providing a new opportunity for the inclusion of Y chromosome markers in sex prediction. In the absence of Y chromosome markers, sex prediction based on X chromosome SNP heterozygosity alone requires careful exclusion of PAR SNPs⁽¹⁾ and the inclusion of only non-pseudoautosomal region (nPAR) SNPs.

The main goals of this study were to identify the PAR region on the X chromosome in sheep using the ARS-UI_Ramb_v2.0 genome assembly and to describe a method for predicting sex in sheep based on the proportion of heterozygous SNPs in the nPAR region of the X chromosome.

2. Materials and Methods

2.1 Training Population

The training dataset consisted of 105 male and 105 female sheep from five breeds (i.e. Corriedale, Creole, Corriedale Pro, Finnish Landrace, and East Friesian) and three crossbreeds (Finnish Landrace × Corriedale, East Friesian × Corriedale, Finnish Landrace × East Friesian), genotyped using the Ovine Infinium® HD SNP BeadChip (606 006 SNPs; Illumina, San Diego, CA, USA). This dataset was used to identify the PAR region on the X chromosome.

2.2 Validation Population

Furthermore, a test dataset consisting of 229 sheep (114 males, 115 females) from three breeds, Corriedale (n=105), Australian Merino (n=67), and Texel (n=57), genotyped using the OvineSNP50 BeadChip (San Diego, CA, USA) was used for validation. Previously, animals with parentage conflicts involving at least one parent were excluded from further analysis due to potential DNA misidentification.

2.3 DNA Extraction

Blood samples were collected through jugular vein puncture from weaned lambs using K_2 EDTA anticoagulant tubes. Genomic DNA was subsequently extracted⁽⁸⁾ from the samples.

2.4 Animal Welfare

Animal handling and blood collection procedures were approved by the INIA Animal Ethics Committee (approval number INIA_2018.2) in accordance with Uruguayan Law 18611.

2.5 Quality Control and Statistical Analyses

Our initial dataset contained 25,883 SNPs situated on the X chromosome based on the ARS-UI_Ramb_v2.0 sheep genome assembly. Genomic data quality control was performed using preGSF90 software⁽⁹⁾. A total of 23,879 SNPs remained after call rate (>0.90) and MAF (>0.01) quality control steps according to Zhan and others⁽²⁾; SNP observed heterozygosity (Ho) was calculated for male and female populations in the training



dataset. The *ggplot2* package from R software⁽¹⁰⁾ was employed to plot the results. The pseudoautosomal boundary (PAB) was established through visual inspection without the use of any software tool, while the PAR region was identified based on the SNPs exhibiting high heterozygosity rates in males. The Y chromosome SNPs were unavailable for analysis since our array did not contain them.

Subsequently, heterozygosity data from the nPAR region was used to determine the sex of animals in a separate dataset containing individuals with recorded sex. The same quality control criteria from the training data were applied to this population, which led to a final set of 1,103 SNPs. The Irish Cattle Breeding Federation (ICBF) provides sex prediction guidelines based on X chromosome nPAR SNPs heterozygosity rates⁽¹⁾: a rate of \leq 5% indicates a male, \geq 15% indicates a female, and rates between 5% and 15% are considered ambiguous for sex determination.

The R *caret* package⁽¹¹⁾ was used to generate a contingency table to evaluate sex prediction accuracy, while the R *pwr* package⁽¹²⁾ was used to conduct a chi-square post hoc power analysis to assess sample size for detecting recorded versus predicted sex differences. Cohen's w was used to quantify the effect size.

3. Results and Discussion

3.1 PAR Region Determination

The boundary separating the PAR and nPAR regions on the X chromosome (PAB) was identified through visual inspection of the scatter plot for males (**Figure 1**a). Males presented an average heterozygosity (Ho) of 0.32 among the first 2009 SNPs (0.00 to 7.24 Mb), while nearly all the remaining 21,870 SNPs (7.62 to 143.12 Mb) showed a Ho close to 0. In contrast, females displayed an average Ho of 0.32 across the entire X chromosome (**Figure 1**b).

Previous studies, using the Oar v4.0 reference genome assembly, reported the sheep PAR region extending from 0 to 7.04 Mb(13). Our results, based on the ARS-UI_Ramb_v2.0 genome assembly, identified this region extending from 0 to 7.24 Mb, which matched the previous research findings. We analysed the X chromosome region from 0 to 7.24 Mb between the ARS-UI_Ramb_v2.0 sequence (NCBI RefSeq assembly: GCF_016772045.1) and the current reference assembly, ARS-UI_Ramb_v3.0 (NCBI RefSeq assembly: GCF_016772045.2), using MAFFT software(14), and found a strong sequence alignment between them (Figure S1 in Supplementary material). The Ovine Infinium® HD SNP BeadChip data indicates that the PAB region extends from the oar3_OARX_9495 SNP (the last PAR SNP at 7.24 Mb) to the oar3_OARX_7679966 SNP (the first nPAR SNP at 7.62 Mb). These findings are crucial because they define two distinct regions on the X chromosome: the PAR region and the nPAR region, with the latter one functioning for sex determination purposes.



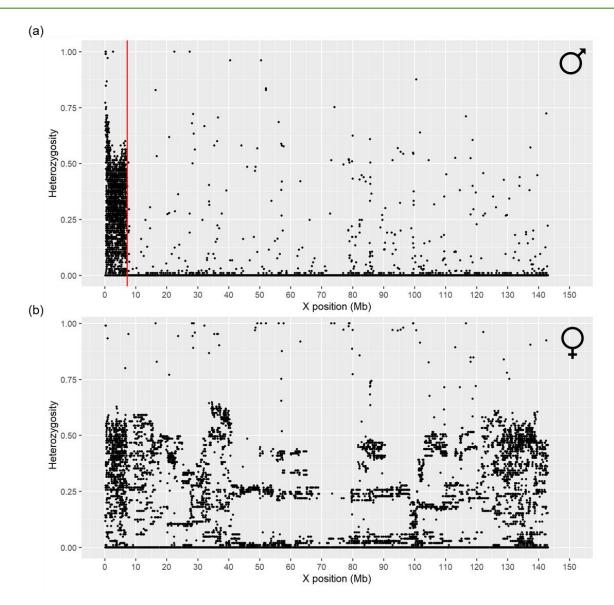


Figure 1. X chromosome SNP heterozygosity distribution among males (a) and females (b) across their respective X chromosome coordinates in the training dataset. The red vertical line in subplot (a) indicates the end of the PAR region at 7.24 Mb

3.2 Sex Prediction

Table 1. Contingency table for recorded and predicted sex for males and females

	PREDICTED_FEMALE	PREDICTED_MALE
Recorded_female	102	13
Recorded_male	1	113

The validation population contained 229 animals, 14 of which had a recorded sex that differed from their predicted sex (**Table 1**), indicating the test achieved 94% accuracy in sex prediction (**Table 2**). The results showed that thirteen recorded females were classified as males, while one recorded male was classified as female, which resulted in precision rates of 99% for males and 89% for females (**Table 2**).



The post hoc power analysis found a Cohen's w effect size of 0.439, indicating a medium effect. The power of the test was calculated to be 0.999 with 1 degree of freedom and a 0.05 significance level, well above the conventional threshold of 0.8. These results suggest that the study is quite powerful and therefore has a high ability to detect real differences if they exist (**Table 2**). Nevertheless, it is important to be careful when generalizing these results to other breeds of sheep. Our validation dataset consisted of Corriedale, Australian Merino, and Texel sheep, which may not entirely capture the breed-specific genomic variation, particularly on the X chromosome. It is advisable to conduct further validation in a larger number of sheep breeds to confirm the robustness of our approach.

Table 2. Classification performance, statistical power, and effect size

CLASSIFICATION PERFORM	ANCE (%)
Accuracy	94
Female misclassification rate	11
Male misclassification rate	1
Precision (female)	89
Precision (male)	99
STATISTICAL POWE	R
Statistical power ($\alpha = 0.05$)	0.999
Cohen's w (effect size)	0.439

A more detailed analysis of the data showed that the recorded male classified as female had a Ho of 0.42 (462 SNPs out of 1,103), and all the recorded females classified as males had a Ho of 0 (0-3 SNPs out of 1,103). Physical examination to confirm the sex of the animals was not possible as none of them are still alive. However, the pedigree file indicated that 12 out of the 13 recorded females classified as males were listed as dams, thus confirming their true female status. On the other hand, the recorded male classified as female was only listed as a lamb in the pedigree file, thus its true male status could not be ascertained. This is most probably due to stud flock management practices where male lambs not selected as sires are culled. Additionally, when this lamb was sampled, the laboratory received two blood tubes with the same identification, which potentially originated from different animals yet only one sample underwent genetic testing. These findings suggest that the recorded sex of these animals is probably correct, and sample mislabelling may have contributed to errors in genotype data.

In addition, the shipment records indicated that four recorded females classified as males and the recorded male classified as female were shipped together in the same 96-well plate, with some of them placed in adjacent wells, which may have caused identification errors. Also, six more females classified as males were shipped together in another 96-well plate, which could suggest a mistake during handling or processing. Since paternity verification was not available for these samples, we cannot state whether the correct samples were genotyped. The other three females classified as males were processed in separate shipments, indicating that biological factors may also be contributing to misclassification rather than sample mislabelling.

Our study achieved a 94% accuracy rate, which represents a significant improvement compared to other sex determination methods such as visual inspection and PCR-based sexing assays⁽¹⁵⁾. Nevertheless, it is lower than the accuracy level observed in other SNP chip array studies⁽²⁾ and studies based on GBS data⁽³⁾. None-



theless, the overall high accuracy obtained indicates that our approach is quite reliable, especially for males, who are classified with very high confidence (99% precision).

Sample mislabelling is a possible reason for some of the errors, but biological factors could also contribute to the 11% female misclassification rate. Sex prediction in females is based on higher observed heterozygosity values (Ho \geq 15%) compared to males (Ho \leq 5%). Since genetic diversity decreases due to inbreeding, some females may not attain the heterozygosity threshold for classification, thus increasing their chances of being misclassified as males or ambiguous sex.

Breed specific differences in X chromosome structure and heterozygosity may also explain discrepancies in female misclassification rates. Merino sheep from stud flocks have been strongly selected for reduced fibre diameter, while animals from an experimental unit have been selected for a combination of traits including fibre diameter, body weight, and clean fleece weight, intensifying directional genetic pressure⁽¹⁶⁾. Although Corriedale is a dual-purpose breed, it has also undergone intensive selection. In this study, Corriedale samples were drawn from registered stud flock animals and from an experimental unit comprising two divergent lines selected for resistance to gastrointestinal parasites⁽¹⁷⁾, both populations showing strong genetic progress. The Texel samples, on the other hand, originated from a central progeny testing unit, which receives rams from stud-breeders for evaluation, but does not implement any selection criteria. These differences in selection history and breeding objectives probably contribute to structural variation in the X chromosome and influence heterozygosity patterns. Consequently, breed-specific variation may affect the heterozygosity threshold used for sex prediction, resulting in different misclassification rates across breeds. Further validation with larger and breed-specific datasets will be important to fully understand the impact of X chromosome variability on SNP-based sex classification.

Though it is recommended to use SNPs from both the X and Y chromosomes for sex prediction, our array did not include Y chromosome SNPs. Previous studies(2)(3)(18) have shown that relying on X chromosome SNPs alone can be problematic in predicting sex due to multiple factors. The genotyping errors that occur because of problems with sample handling can be considered as random errors which may lead to incorrect sex assignment. Also, inbreeding may result in misclassification, especially among highly inbred females who inherit the same intact X chromosome from a common ancestor, which gives them X chromosome profiles similar to those of males(18). Although the average pedigree inbreeding coefficient for Uruguayan sheep is relatively low (1.1%), with Corriedale at 1.35% and Merino at 0.63%, these values are based on our own calculations using registered stud flock animals from genetic evaluations. Reduced genetic variation in more inbred populations may increase the chances of females not meeting the heterozygosity threshold for accurate classification. Sex chromosome aneuploidies, such as XO in females (where there is only one X chromosome) and XXY in males (where there is an extra X chromosome), have been observed in sheep⁽¹⁹⁾⁽²⁰⁾⁽²¹⁾. Even though these conditions are quite rare, they involve abnormalities in the number of sex chromosomes, which may result in atypical X chromosome profiles. Although we did not directly analyse sex chromosome aneuploidies in our dataset, a study using a sample of 146,431 juvenile cattle reported the frequencies of XO in females at 0.0048% and XXY in males at 0.0870%⁽²²⁾. Future research including SNP genotype intensity data may be used to determine the frequency of such aneuploidies in Uruguayan sheep. Intersex conditions such as XX sex reversal and androgen insensitivity syndrome, although rare, also pose a challenge. In XX sex reversal, the individuals have an XX karyotype (XX) but have male gonadal and/or phenotypic sex(23), whereas in androgen insensitivity syndrome, the individuals have an XY genotype but have hormonally active but retained testes and develop female external genitalia due to defective androgen receptors (24). These rare anomalies can result in chromosomal-phenotypic sex discordance, which may decrease the reliability of SNP-based classification. Importantly, these genetic and physiological factors are not independent but can interact, as inbreeding may increase the incidence of sex chromosome abnormalities and intersex conditions, which will make classification more



difficult. Together, these limitations highlight the need to use markers from both the X and Y chromosomes to improve the accuracy of sex prediction methods.

The sex prediction method evaluated in this research study demonstrates practical value for commercial sheep genotyping pipelines. The test provides an accurate way to predict animal sex when Y chromosome information is missing and simultaneously helps detect animals with aneuploidies or intersex conditions. Genetic analysis reliability and breeding programs effectiveness would rise because misclassification risks would decrease through this approach.

The integration of sequencing data with SNP-based sex prediction represents a promising direction for future research. SNP arrays represent a cost-effective solution, but sequencing technologies deliver enhanced accuracy. These technologies allow the direct analysis of sex chromosome sequences, which leads to better identification of genetic anomalies. This approach would help validate results and identify errors while designing better SNP marker panels. An example is the recent high-quality Y chromosome assembly for sheep developed by Olagunju and others⁽⁷⁾, that shows promise for integrating Y chromosome markers in sex prediction.

4. Conclusion

We determined the location of the PAR region on the sheep X chromosome using the ARS-UI_Ramb_v2.0 genome assembly and described a method which uses X chromosome SNPs to predict sex. This method generates precise sex predictions across various sheep breeds, making it an effective quality control tool in the absence of Y chromosome data. The method demonstrated positive results but requires further research using sequencing data along with studies on genetic anomalies, like aneuploidies and intersex conditions, to achieve better validation and refinement.

Acknowledgements

Funding for data used in this project was provided by FAO project "Caracterización productiva y conservación de ovinos criollos del Uruguay", the National Agency for Research and Innovation (ANII, Uruguay) - Innovagro (FSA_1_2013_1_12560), and the National Agricultural Research Institute (INIA_CL_20 and INIA CL_38).

This research was conducted as part of a PhD thesis at the School of Agronomy, University of the Republic, Uruguay.

Transparency of data

Data not available: The data set that supports the results of this study is not publicly available.



Author contribution statement

	B Carracelas	EA Navajas	G Ciappesoni	l Aguilar
Conceptualization				
Data curation				
Formal analysis				
Investigation				
Methodology				
Supervision				
Visualization				
Writing – original draft				
Writing – review and editing				

References

- (1) McClure MC, McCarthy J, Flynn P, McClure JC, Dair E, O'Connell DK, Kearney JF. SNP data quality control in a national beef and dairy cattle system and highly accurate SNP based parentage verification and identification. Front Genet. 2018;9:84. Doi: 10.3389/fgene.2018.00084.
- ⁽²⁾ Zhang I, Couldrey C, Sherlock R. Using genomic information to predict sex in dairy cattle. Proc N Z Soc Anim Prod. 2016;76:26-30.
- (3) Bilton TP, Chappell AJ, Clarke SM, Brauning R, Dodds KG, McEwan JC, Rowe SJ. Using genotyping-by-sequencing to predict gender in animals. Anim Genet. 2019;50(3):307-10. Doi: 10.1111/age.12782.
- (4) Raudsepp T, Das PJ, Avila F, Chowdhary BP. The pseudoautosomal region and sex chromosome aneuploidies in domestic species. Sex Dev. 2012;6(1-3):72-83. Doi: 10.1159/000330627.
- (5) Johnson T, Keehan M, Harland C, Lopdell T, Spelman RJ, Davis SR, Rosen BD, Smith TPL, Couldrey C. Identification of the pseudoautosomal region in the Hereford bovine reference genome assembly ARS-UCD1.2. J Dairy Sci. 2019;102(4):3254-8. Doi: 10.3168/jds.2018-15638.
- (6) Liu R, Low WY, Tearle R, Koren S, Ghurye J, Rhie A, Phillippy AM, Rosen BD, Bickhart DM, Smith TPL, Hiendleder S, Williams JL. New insights into mammalian sex chromosome structure and evolution using high-quality sequences from bovine X and Y chromosomes. BMC Genomics. 2019;20(1):1000. Doi: 10.1186/s12864-019-6364-z.
- (7) Olagunju TA, Rosen BD, Neibergs HL, Becker GM, Davenport KM, Elsik CG, Hadfield TS, Koren S, Kuhn KL, Rhie A, Shira KA, Skibiel AL, Stegemiller MR, Thorne JW, Villamediana P, Cockett NE, Murdoch BM, Smith TPL. Telomere-to-telomere assemblies of cattle and sheep Y-chromosomes uncover divergent structure and gene content. Nat Commun. 2024;15(1):8277. Doi: 10.1038/s41467-024-52384-5.
- (8) Medrano JF, Aasen E, Sharrow L. DNA extraction from nucleated red blood cells. Biotechniques. 1990;8(1):43.
- (9) Aguilar I, Misztal I, Tsuruta S, Legarra A, Wang H. PREGSF90 POSTGSF90: computational tools for the implementation of single-step genomic selection and genome-wide association with ungenotyped individuals in BLUPF90 programs. In: Proceedings 10th World Congress of Genetics Applied to Livestock Production. Vancouver: American Society of Animal Science; 2014. p. 680.
- (10) Wickham H. ggplot2: elegant graphics for data analysis [Internet]. [place unknown]: The tidyverse team; [cited 2025 May 27]. Available from: https://ggplot2.tidyverse.org



- (11) Kuhn M. Building Predictive Models in R Using the caret Package. J Stat Softw. 2008;28(5):1-26. Doi: 10.18637/jss.v028.i05
- (12) Champely S. pwr: basic functions for power analysis [Internet]. Version 1.3-0. 2020 [cited 2025 May 27]. Available from: https://CRAN.R-project.org/package=pwr
- (13) Shihabi M, Lukic B, Cubric-Curik V, Brajkovic V, Oršanić M, Ugarković D, Vostry L, Curik I. Identification of selection signals on the X-Chromosome in East Adriatic Sheep: a new complementary approach. Front Genet. 2022;13:887582. Doi: 10.3389/fgene.2022.887582.
- (14) Katoh K, Rozewicki J, Yamada KD. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Brief Bioinform. 2019;20(4):1160-6. Doi: 10.1093/bib/bbx108.
- (15) Ristanic M, Stanisic L, Maletic M, Glavinic U, Draskovic V, Aleksic N, Stanimirovic Z. Bovine foetal sex determination-Different DNA extraction and amplification approaches for efficient livestock production. Reprod Domest Anim. 2018;53(4):947-54. Doi: 10.1111/rda.13193.
- (16) Ramos Z, Garrick DJ, Blair HT, De Barbieri I, Ciappesoni G, Montossi F, Kenyon PR. Genetic trends for production and reproduction traits in ultrafine Merino sheep of Uruguay. J Anim Breed Genet. Forhcoming 2025. Doi: 10.1111/jbg.12937.
- (17) Ciappesoni G, Marques C, Navajas EA, Peraza P, Carracelas B, Vera B, Van Lier E, De Barbieri I, Salada S, Monzalvo C, Castells D. Breeding for sheep parasite resistance in extensive production systems in Uruguay: from phenotype to genotype. In: Viljoen G, Garcia Podesta M, Boettcher P, editors. International Symposium on Sustainable Animal Production and Health Current status and way forward. Rome: FAO; 2023. p. 224-36.
- (18) Berry DP, Spangler ML. Animal board invited review: practical applications of genomic information in livestock. Animal. 2023;17(11):100996. Doi: 10.1016/j.animal.2023.100996.
- (19) Bruere AN, Marshall RB, Ward DP. Testicular hypoplasia and XXY sex chromosome complement in two rams: the ovine counterpart of Klinefelter's syndrome in man. J Reprod Fertil. 1969;19(1):103-8. Doi: 10.1530/jrf.0.0190103.
- ⁽²⁰⁾ Zartman DL, Hinesley LL, Gnatkowski MW. A 53,X female sheep (Ovis aries). Cytogenet Cell Genet. 1981;30(1):54-8. Doi: 10.1159/000131589.
- (21) Baylis MS, Wayte DM, Owen JB. An XO/XX mosaic sheep with associated gonadal dysgenesis. Res Vet Sci. 1984;36(1):125-6.
- (22) Ryan CA, Purfield DC, Matthews D, Canedo-Ribeiro C, Valldecabres A, Berry DP. Prevalence of sexchromosome aneuploidy estimated using SNP genotype intensity information in a large population of juvenile dairy and beef cattle. J Anim Breed Genet. 2024;141(5):571-85. Doi: 10.1111/jbg.12866.
- (23) Raudsepp T, Chowdhary BP. Chromosome aberrations and fertility disorders in domestic animals. Annu Rev Anim Biosci. 2016;4:15-43. Doi: 10.1146/annurev-animal-021815-111239.
- (24) Berkowic Y, Edwards LA, Busch R, Pérez-Nogués M, Stevens SE, Booth M, McNabb BR. Male pseudohermaphrodite in a Shetland cross sheep: a case report. Clinical Theriogenology. 2017;9(4):605-13.



Supplementary Material

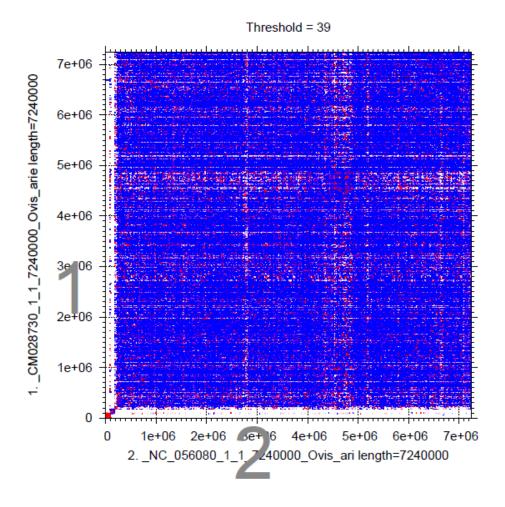


Figure S1. Sequence comparison of the X chromosome region (0 to 7.24 Mb) between the ARS-UI_Ramb_v2.0 sequence (NCBI Ref-Seq assembly: GCF_016772045.1) and the current reference assembly, ARS-UI_Ramb_v3.0 (NCBI Ref-Seq assembly: GCF_016772045.2)